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### A. INTRODUCTION

#### 1. Theory

This method exploits the weakly basic character of Macrolide and lincosamide (mac/linc) antibiotics. Sample tissue is made basic and extracted with ethyl acetate. Analytes are then partitioned into an acidic buffer and further cleaned up by extraction of the buffer solution with an organic solvent. The buffer is then made basic and analytes are re-extracted into ethyl acetate, evaporated to dryness, redissolved in mobile phase, and analyzed by ion trap HPLC/MS/MS. Confirmation is based on comparison of sample and MS/MS spectral data with that of a fortified tissue standard or external standard.

### 2. Applicability

The method is applicable to liver, kidney and muscle tissues in beef, pork, and poultry. Macrolides detectable to a level of 0.1 ppm by this method are Lincomycin, Clindamycin, Erythromycin, Tylosin, Tilmicosin, and Pirlimycin.

#### B. EQUIPMENT

Equivalent apparatus may be substituted for those listed below.

### 1. Apparatus

- a. Centrifuge IEC-HN-S11.
- b. Waring Blender Cat. No. 33BL79 equipped with a 40 mL blending jar.
- Vortex mixer Genie 2, Fisher Scientific.
- d. pH meter Cat. No. 215 with a Accumet micro combination electrode, Cat. No.13-620-95, Denver Instrument Co.
- e. pH meter Cat. No 370 with ATC probe, Orion.
- f. Balance accurate to 0.0001 g, Cat. No. MT5, Mettler.
- g. Nitrogen evaporator TurboVap LV, Zymark.
- h. Volumetric flasks 100 mL and 10 mL class A volumetric flasks.
- i. 15 mL glass centrifuge tubes Cat. No. 73790, Kimble for Zymark TurboVap.
- j. 15 mL glass centrifuge tubes Cat. No. 73785, Kimble.
- k. Autosampler vials 1.8 mL wide mouth glass, Cat. No. 5182-0543, Agilent.
- I. Micropipetters covering the range from  $50\mu$ L  $5000 \mu$ L.
- m. Pasteur pipettes 5 3/4 in Cat. No. 53283-910; Lab Depot.

### 2. Instrumentation

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- Ion trap mass spectrometer Finnigan LCQ-deca equipped with an APCI LC interface and LCQ Xcalibur data system, or equivalent.
- b. LC system Quaternary pump equipped with degassing capability and autosampler. Thermo-Finnigan Surveyor HPLC and autosampler.
- c. LC column. Zorbax SB-C  $_{18}$  2.1 x 150 mm containing 5  $\mu$ m particles, preceded by a 0.2  $\mu$ m frit filter.

### C. REAGENTS AND SOLUTIONS

Equivalent reagents and solutions may be substituted for the following unless otherwise indicated:

### 1. Reagents

- a. Methanol (MeOH), LC grade Mallinckrodt UltimAR grade. (Do not substitute.)
- Water, LC grade House distilled water passed through Waters MilliQ deionization system.
- c. Acetonitrile UV grade, Cat. No. 015-4, Burdick & Jackson.
- d. Phosphoric acid ACS grade, Cat. No. P6560, Sigma.
- e. Ethyl acetate UltimAr grade, Cat. No. U-553-10, Mallinckrodt.
- f. Hexane Cat. No. HX0296-1, EM OmniSolv.
- g. Potassium phosphate monobasic (KH<sub>2</sub>PO<sub>4</sub>) HPLC grade, Cat. No. P286-1, Fisher.
- h. Formic acid ACS grade, Cat. No. F-4636, Sigma.
- i. Potassium carbonate ACS grade, Cat. No. P6037, Sigma.

#### 2. Solutions

Note: Solutions may be stored at room temperature unless otherwise noted.

- a. 50:50 (v/v) Methanol/water:
  - Mix 50 mL methanol with 50 mL of water. Stable for 6 months.
- b. 5:95 (v/v) acetonitrile/water + 0.1% formic acid:
  - Mix 50 mL acetonitrile, 950 mL water and 1.0 mL formic acid in a 1 liter graduated cylinder. Stable for 6 months.
- c. 95:5 (v/v) acetonitrile/water + 0.1% formic acid:
  - Mix 950 mL acetonitrile, 50 mL water and 1.0 mL formic acid in a 1 liter graduated cylinder. Stable for 6 months.
- d. 0.2 M potassium phosphate monobasic, pH 4.00:

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Weigh 13.6 g of KH<sub>2</sub>PO<sub>4</sub> into a 500 mL volumetric flask and dilute to volume with water. Adjust the pH to 4.00 with 1:20 phosphoric acid. Store Refrigerated. Stable for 3 months. Warm to room temperature before use.

e. 1:20 (v/v) phosphoric acid/water:

Dilute 5 mL of concentrated phosphoric acid to 100 mL with water in a volumetric flask.

f. 1:1 (v/v) ethyl acetate:hexane:

Mix equal volumes of ethyl acetate and hexane.

g. 2 M potassium carbonate:

Weigh 27.64 g of K<sub>2</sub>CO<sub>3</sub> into a 100 mL volumetric flask and dilute to mark with water.

#### D. STANDARDS

#### Names and Sources

Name	Cat. No.	Source
Lincomycin	L6004	Sigma
hydrochloride		
Clindamycin	C5269	Sigma
hydrochloride		
Erythromycin	E0774	Sigma-USP
Tylosin tartrate	T6134	Sigma
Tilmicosin		Lily research Labs
Pirlimycin		Pfizer Corp.
hydrochloride		

### 2. Preparation of Standard Solutions

Note: Equivalent standards and solutions may be substituted for any of the following.

a. Individual drug stock standard solutions (100 µg/mL):

Using vendor's stated purity, or water and salt content, calculate the amount of material which contains 10 mg drug base. Weigh out approximately this amount, accurately recording weight to nearest 0.1 mg. Transfer to 100 mL glass volumetric flask and dilute to volume with methanol. Calculate exact concentration based on purity and actual weight. Stable for 6 months at <-10°C.

b. Mixed standard fortification solution (5 μg/mL):

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Add 500  $\mu$ L of each of the above drug stock solutions to a 10 mL volumetric flask and dilute to volume with methanol. Stable for 6 months at <-10  $^{\circ}$ C.

#### E. SAMPLE PREPARATION AND CLEANUP

- 1. Blend sufficient whole liver, muscle or kidney tissue for each sample in a 40 mL. Blending jar. A separate sample holding/receiving section may homogenize sample using their equipment.
  - Note: After each blending and weighing, rinse the blending jar with hot water, deionized water, methanol, hexane and finally with methanol.
- Weigh 5.00 ± 0.10 g tissue into a 50 mL disposable polypropylene centrifuge tube. If muscle tissue is to be analyzed, add approximately 3.0 mL of water and mix well with a microspatula.

Note: Prepare Controls (to be included as part of each sample batch) at this time:

- a. Negative controls are tissues from animals known to be free of drugs. If these are not available, tissue from an unknown source may be used provided it is first tested and shown to be free of contaminants. Store tissue frozen, preferably at
  < - 10 °C prior to analysis.</li>
- b. Positive controls are negative tissues that have been fortified with mac/lincs before extraction. To prepare a 0.1 ppm fortified sample, add 100  $\mu$ L (5  $\mu$ g/mL) mixed standard solution (D.3.b. above), then vortex 10 to 20 seconds (If muscle tissue, stir with a microspatula instead).
- 3. Prepare an aqueous tissue suspension having a pH of 9.8-10.2 by adding aliquots of  $2M K_2CO_3$  and mixing until a stable pH in that range is obtained (usually requires  $300-500 \mu L$  of  $2M K_2CO_3$ ). Vortex or stir with a microspatula (muscle) after each addition. Measure pH using a micro pH electrode just touching the surface of the solution.
- 4. Add 30 mL of ethyl acetate cap and shake 3 min. Centrifuge for 10 min at approximately 2000 rpm (approximately 800 g rcf). Pour the supernatant into a second 50 mL polypropylene centrifuge tube.
- 5. Repeat step 4 above with 15 mL of ethyl acetate and add to the polypropylene centrifuge tube. Remove any oily residue at the bottom of the tube of the combined ethyl acetate extracts with a Pasteur pipette (occasionally seen with muscle samples).
- 6. To the approximately 45 mL of combined organic phase, add 2.0 mL of 0.2M KH<sub>2</sub>PO<sub>4</sub>. Shake 3 min. and centrifuge at approximately 2000 rpm for 6 min. Transfer the bottom aqueous layer with a Pasteur pipette and bulb to a 15 mL glass centrifuge tube.
- 7. Repeat step 6 twice. Retain each sample's Pasteur pipette and use for all three transfers and then discard.
- 8. To the approximately 6 mL of combined aqueous solution, add 5 mL of a 1:1 ethyl acetate:hexane solution and invert the tube to mix 10 times. Centrifuge at 2000 rpm for

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4 min. Discard the top organic layer.

Note: Be sure to remove the entire top organic layer.

- Adjust the pH of the aqueous solution to 9.8-10.2 by adding approximately 900 to 1200 μL of 2M K<sub>2</sub>CO<sub>3</sub> (check the pH using a micro pH electrode just touching the surface of the solution)
- 10. Add 4.0 mL of ethyl acetate to the aqueous solution and shake 3 min. Centrifuge at 2000 rpm for 4 min. Transfer the upper organic layer to a 15 mL disposable glass centrifuge tube (Zymark tube) with a Pasteur pipette.
- 11. Repeat step 10 twice. Retain each sample's Pasteur pipette and use for all three transfers and then discard.
- 12. Evaporate the combined organic solution to near dryness (approximately 200 μL) in a TurboVap maintained at approximately 40 °C. Take the remainder of the solution to dryness at room temperature.
- 13. Dissolve the residue in 500  $\mu$ L. of 50:50 (v/v) methanol/water. Mix for a total of 30 seconds prior to filtering through a 0.2  $\mu$ m PTFE syringe filter into a 1.8 mL autosampler vial.

### F. ANALYTICAL PROCEDURE

Note – Instrumental parameters yielding equivalent analytical results may also be used.

- 1. Instrument Operating Parameters LC System
  - a. Install and degas mobile phases and install column and guard cartridge per manufacturer's instructions. Set initial composition to flow 5/95 acetonitrile/water+0.1% formic acid at 300 µL/min.
  - b. Set-up the HPLC to run the following linear gradients

Time in min.	Flow in mL/min.	A (5/95 A/W+0.1%fa)	B (95/5 A/W+0.1%fa)
0.00	0.30	100%	0%
15.00	0.30	0%	100%
15.10	0.30	100%	0%
25.00	0.30	100%	0%

- c. Set injection volume to 20 μL.
- d. Use a needle wash step with MeOH.

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- 2. Instrument Operating Parameters Mass Spectrometer
  - a. Mass Spectrometer. Calibrate the Finnigan LCQ ion trap mass spectrometer with electrospray interface according to the manufacturer's specifications.
  - b. Set Capillary Temp to 150 °C.
  - c. Operate in Pos mode.
  - d. Flow inject the external standard through a 5 μL loop and obtain the precursor ion centroids. The following settings resulted in optimal ion intensities:

Scan range for data dependant aq.	400-950 d
Capillary temperature	150 °C
APCI vaporizer temperature	450 °C
Sheath gas flow	60
Aux gas flow	5
Capillary voltage	10v
Tube lens offset	2v
Micro scans	1
Ion time	100 msec.
Source current	5.00 µA

- 3. Procedure for Instrumental Analysis of Samples, Controls and Standards
  - a. Turn on pump and set up mass spectrometer. Equilibrate column in mobile phase at 0.30 mL/min for at least 30 min.
  - b. Flow inject the external 100 ppb standard through a 5  $\mu$ L loop and obtain the precursor centroids.

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c. Inject the external standard through the HPLC system and acquire spectra using data dependant scanning under the following procedure:

	_	
Scan	Event	Details
1	Pos	Full scan 400-950 D
2	Pos	Dep MS <sup>2</sup> most intense
		ion from parent list (1)
3	Pos	Dep MS <sup>2</sup> 2nd most
		intense ion from
		parent list (1)
Dependent Data Settings		
Parent mass list	407.1, 411.1, 425.1,	There maybe slight
	688.1, 734.2, 869.5,	changes during
	916.3	optimization.
2. Charge state	1	
3. Isolation width	2.00	
4. Activation amplitude	34.0	
5. Activation Q	0.250	
6. Activation time	30.00	
7. Min. signal required	100000	
8. Min. MS <sup>n</sup> signal required	5000	

- d. Inject the recovered standard and verify retention time, and divert valve switching time.
- e. Inject the negative control and the sample extracts. If necessary to control carryover, precede each sample analysis with a sample diluent injection.
- f. Column, Pump, and APCI Interface Care. At the end of set of analyses, flush the column for 30 min with acetonitrile + water (60 + 40) at 0.30 mL/min.

#### G. CONFIRMATION

1. Data Processing.

Use the QUAL Browser to view total ion current, base ion chromatogram, and/or a reconstructed ion chromatogram for each drug for each data file. Note retention time of any visible peaks in a drug window. Generate averaged spectra across the retention time window for each drug. This is usually from near the start to near the end of the peak visible in the chromatograms, though a smaller range may be used to avoid a spurious ion spike. Where no peak is visible, use the same settings as in a contemporaneous fortified or positive control extract.

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### 2. Confirmation Criteria.

- a. Retention times of extract peaks must match the peak retention time in a contemporaneous (within same analysis set on same day) fortified control extract chromatogram within 0.2 min.
- b. The mac/linc peak in the total ion chromatogram (TIC) is present at an S/N ratio of at least 3/1. This is estimated by visual inspection of the TIC.
- c. The spectra from the extract must visually match spectra from external standards in the same data set. The base ion must be the same. The base ion, two qualifying ions and at least an additional product ion shall be present and readily distinguished from background and matrix ions. There should be a general absence of nonspecific ions. Major specific ions for each mac/linc are listed below:

Mac/linc	Approx. retention time (min)	Precursor ion	Spectra Range	Base Product ion	Product Ions
Lincomycin	4.75	407.1	100-420	359	126*,172, 389*
Pirlimycin	6.71	411.1	100-425	363	327*, 375, 393*
Clindamycin	7.10	425.1	105-440	377	172, 126*, 389*, 407
Tilmicosin.	7.56	869.5	225-880	696	522*, 678*, 738
Erythromycin	8.33	734.2	190-745	576	558*, 716, 522*, 698, 540
Tylosin	8.68	916.3	240-930	772	407*, 598*, 754

<sup>\*</sup> Qualifying ions.

- d. The quality assurance positive and negative control samples confirm and fail to confirm, respectively, for the presence of the appropriate drug.
- 3. Criteria for Repeating an Analysis.

Sample analyses may be repeated under the following conditions:

a. The conditions described in G.2.d are not met.

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- b. The instrument is suspected to be malfunctioning, as demonstrated by: clearly aberrant standard spectra; failure of a calibration check performed shortly after analysis of the sample set; instrumental parameters, especially vacuum readings, outside of normal operating range; or other conditions noted and documented by the analyst.
- c. There is suspected carryover from a previous high concentration sample or standard. In this case, the sample should be reanalyzed after the cause of the carryover has been identified and measures taken to prevent its recurrence.
- d. There is strong evidence of mac/linc presence, but multiple extraneous ions with relative abundance exceeding that of mac/linc's base ion prevent unambiguous confirmation. In this case, it may be appropriate to reanalyze the suspected positive sample together with a chromatographic standard, and negative and positive QA controls.

### H. HAZARD ANALYSIS

- 1. Required Protective Equipment
  - a. Personal protective equipment (PPE) includes gloves, safety glasses, and lab coat, where applicable.
  - b. Fume hood.

### 2. Hazards

Procedure Step	Hazard	Recommended Safe Procedures
Mac/Linc antibiotic Standards	Can cause kidney damage.	Wear PPE when handling standards.
Methanol	Highly flammable, and may	Use under a fume hood away
Hexane	produce toxic effects to skin,	from all electric devices and open flames. Avoid breathing
Ethyl acetate	eyes and the respiratory system.	vapors.
Acetonitrile	Highly flammable and toxic liquid. May cause skin irritation.	Use under a fume hood away from all electric devices and open flames. Treat as cyanide. Avoid breathing vapors
Formic acid and Phosphoric acid	Corrosive. Danger of chemical burns.	Prepare solutions in a fume hood. Wear PPE, and avoid contact with skin.

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### 3. Disposal Procedures

Procedure Step	Hazard	Recommended Safe Procedures
Methanol/acetonitrile Hexane, Ethyl acetate	See section 2 above	Collect waste in a sealed container and store in a cool, well ventilated, flammable liquid storage area/cabinet for disposal in accordance with local, state, and Federal regulations.
Acids and acidic reagents.	See section 2 above	Collect waste in a sealed container and store in a cool, well ventilated, acid liquid storage area/cabinet for disposal in accordance with local, state, and Federal regulations.

### I. QUALITY ASSURANCE PLAN

1. Performance Standard

Refer to Section G.2 for Confirmation Criteria.

- 2. Readiness to Perform (FSIS Training Plan)
  - a. Familiarization
    - i. Phase I: Standards Inject external standard solutions (D.2.c) in duplicate on at least three different days, and verify instrument response is adequate for confirmatory purposes.
    - ii. Phase II: Fortified samples Analyze one fortified bovine kidney, liver, and muscle and one blank bovine kidney, liver, and muscle. On a subsequent separate day analyze one fortified porcine kidney, liver, and muscle and one blank porcine kidney. liver, and muscle. On a subsequent separate day analyze one fortified poultry kidney, liver, and muscle and one blank poultry kidney, liver, and muscle. (total 18 samples)
      - Note: Phase I and Phase II may be performed concurrently.
    - iii. Phase III: Check samples for analyst accreditation.
  - b. 6 check samples fortified at levels between 1–2 times MPL using analytes and concentrations unknown to the analyst. These six unknowns shall use two bovine kidney, two bovine liver, and two bovine muscle tissues. Each set must

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include a positive control and a negative control. Approval from the Supervisor and the Laboratory Quality Assurance Manager (QAM) is required to commence official analysis.

c. Acceptability criteria.

Refer to section I.1 above

- 3. Intralaboratory Check Samples
  - a. System, minimum contents.
    - i. Frequency: One per week per analyst when samples analyzed.
    - ii. Records are to be maintained for review.
  - b. Acceptability criteria.

If unacceptable values are obtained, then:

- i. Stop all official analyses by that analyst.
- ii. Take corrective action.
- 4. Sample Acceptability and Stability
  - a. Matrices: Bovine, porcine and poultry liver, muscle, and kidney.
  - b. Sample receipt, minimum weight: approximately 50 grams.
  - c. Condition upon receipt: chilled or frozen.
  - d. Sample storage:
    - i. Time: 2 weeks for blended/ homogenized samples
    - ii. Condition: frozen (less than -10 °C)
- 5. Sample Set Each sample set must include the following:
  - a. Negative kidney control sample and a positive kidney control sample.
  - b. Samples.
- 6. Sensitivity
  - Method has been demonstrated to function reliably at MPLs specified below.

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### b. Minimum proficiency levels<sup>1</sup> (MPL), in ppm, by tissue.

	<u>Liver</u>	<u>Kidney</u>	<u>Muscle</u>
lincomycin	0.1	0.1	0.1
pirlimycin	0.1	0.1	0.1
clindamycin	0.1	0.1	0.1
Tilmicosin	0.1	0.1	0.1
erythromycin	0.1	0.1	0.1
tylosin	0.1	0.1	0.1

<sup>&</sup>lt;sup>1</sup>Minimum analyte concentration analyst is expected to reliably confirm.

### J. WORKSHEET

Example of a worksheet, on the following pages, may be copied for use.

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### MACROLIDE/LINCOSAMIDE CONFIRMATION WORKSHEET

Analyst:	Instrument used:	
,	Method File Name:	
Date Started:		
	Mixed Fort. Std.:	
Date Completed:		
	A: 5/95 ACN/H <sub>2</sub> O + 0.1%FA:	
Set Number:		
Species:		
Reviewed By Init/Date:	-	

						Macrolide/Lincosamide		Macrolide/Lincosamide				Macrolide/Lincosamide								
Smp #	Lab No.	Form #	Tissue Type	Smp wt (5 ± 0.10 g).	Smp pH (8.9 –10.2)	Ret Time (min)	Ret. Time within ± 0.20 min of Fort Extract(s)	RIC>3x Noise Level?	Qualifying/ Base Ion(s) Present?	Visual Match?	Ret Time (min)	Ret. Time within ± 0.20 min of Fort Extract(s)	RIC>3x Noise Level?	Qualifying/ Base Ion(s) Present	Visual Match ?	Ret. Time (min)	Ret. Time within ± 0.20 min of Fort Extract(s)	RIC>3 x Noise Level?	Qualifying/ Base Ion(s) Present?	Visual Match ?
																				<del>                                     </del>
																				<del></del>

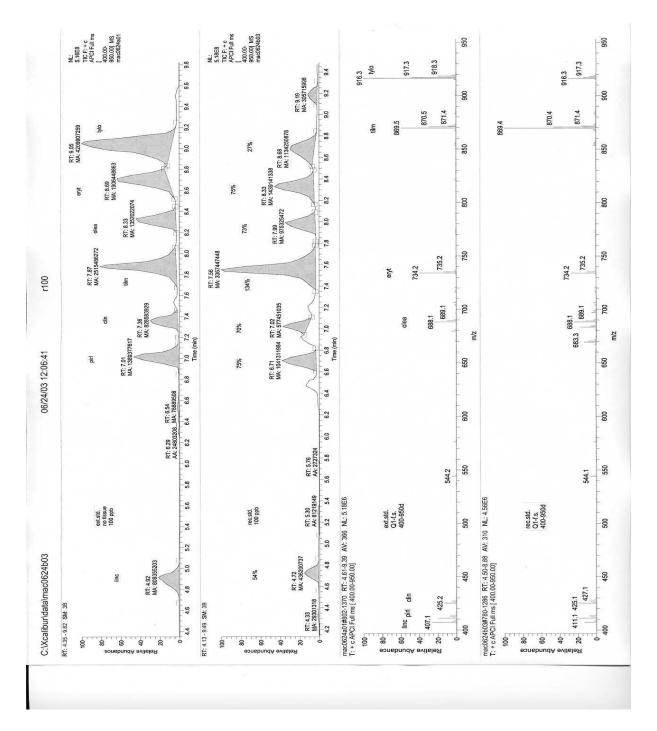
Note: Place a check mark where applicable to indicate a positive response. A check mark in each of the four columns indicates confirmation.

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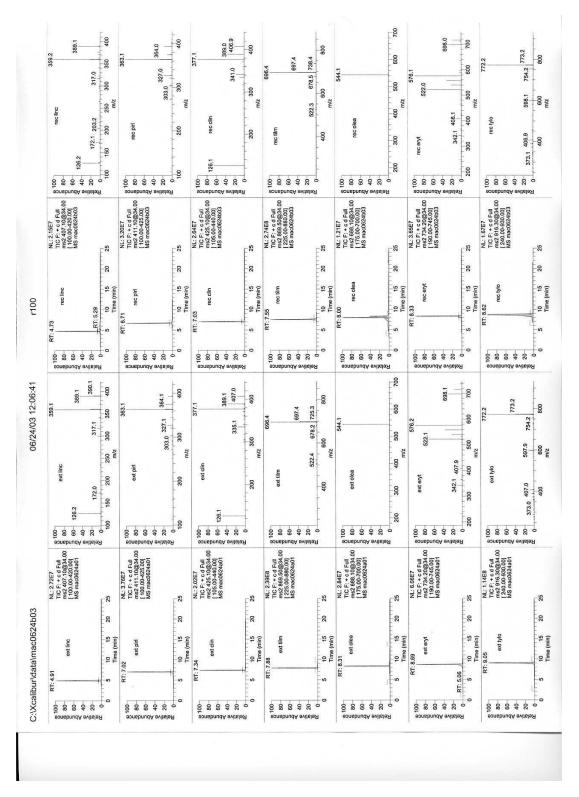
### K. APPENDIX

Retention times and spectra for Macrolide/Lincosamide Antibiotics are shown in the following three pages.

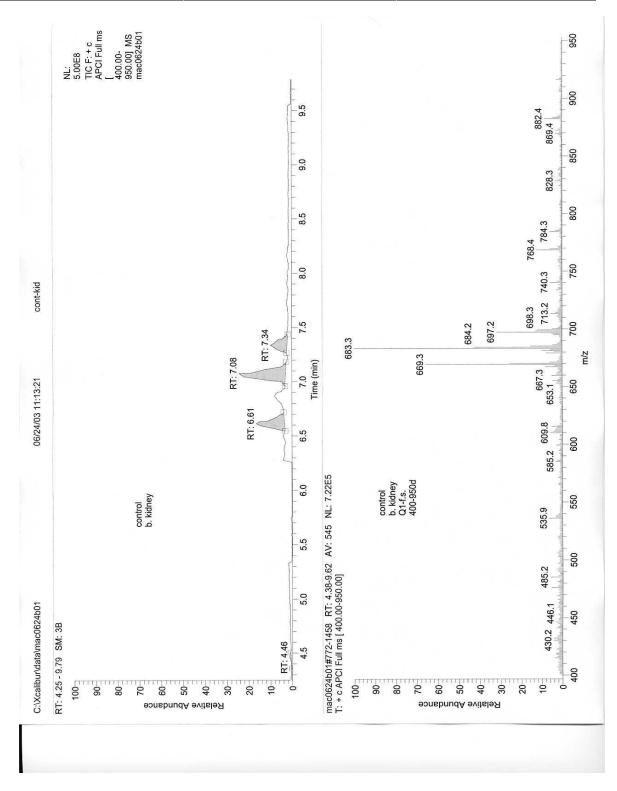
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### **Approvals**

Approved by:	Date Approved
Tom Mallinson	11/13/03
Terry Dutko	11/7/03
Jess Rajan	11/12/03
Charles Pixley	11/7/03
Phyllis Sparling	11/12/03